

USE OF HETEROPOLYACIDS FOR TLC ANALYSIS OF TRITERPENE GLYCOSIDES

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UDC 543.544:547.918

We reported earlier on reagents for detecting triterpene glycosides on TLC plates that included various salts or aromatic aldehydes and H₂SO₄ [1]. In this communication we describe the use for these purposes of mixtures containing the heteropolyacids phosphotungstic H₇[P(W₂O₇)₆] (**1**), phosphomolybdic H₇[P(Mo₂O₇)₆] (**2**), and silicotungstic H₈[Si(W₂O₇)₆] (**3**), which also give color reactions with triterpene glycosides.

Acid **1** is used as an alcoholic solution (25%) to detect triterpene glycosides on TLC plates because it gives distinct and well preserved spots [2]. Acid **2** is a general reagent and is used to identify not only saponins but also lipids, antioxidants, and other compounds [3]. Acid **3** has been used to detect vitamin D on TLC plates and paper [4] and as a qualitative reagent for alkaloids [5]. The use of the last to detect triterpene glycosides has not been reported.

We used solutions of **1-3** in CHCl₃:CH₃OH and with added *p*-hydroxybenzaldehyde or H₂SO₄. Oleanane-type glycosides with hederagenin and oleanolic and echinocystic acids as the aglycons were detected. It has been found that treatment of the chromatograms with CHCl₃:CH₃OH solutions of **1** and **3** with subsequent heating causes spots of hederagenin and its glycosides to acquire a bluish-violet color; oleanolic and echinocystic acids and their glycosides, reddish-pink. On the other hand, **2** gives in all instances blue spots on a yellowish-green background. Thus, this reagent cannot initially differentiate glycosides by the type of aglycon.

A solution with a high concentration of **1** (25%) [2] gives a rather strong background if the chromatograms are heated. Therefore, we used a lower concentration of **1**. After the chromatograms were treated with this reagent and heated, the spots of the compounds appeared on a light pinkish-brown background. On the other hand, there was practically no visible background if **3** was used in this instance.

If chromatograms were treated with reagents with added *p*-hydroxybenzaldehyde or H₂SO₄, the same color effects were noted for **1** and **3** as when the CHCl₃:CH₃OH solutions were used. However, in this instance the spots were more distinct, more strongly colored, and more stable. Spots of glycosides persisted longer than any if treated with reagents containing H₂SO₄. Furthermore, adding *p*-hydroxybenzaldehyde and H₂SO₄ increased the sensitivity of the detecting reagents and enabled the concentration of heteropolyacid to be decreased. Detection of glycosides and pure aglycons using **2** with added *p*-hydroxybenzaldehyde or H₂SO₄ also did not enable the type of aglycon to be established initially because all spots were dark blue on a blue background.

Triterpene glycosides of known structure that were isolated by us from various organs of Crimea ivy *Hedera taurica* Carr. and *H. canariensis* Willd. were used for TLC. These were oleanolic acid 3-*O*- α -L-arabinopyranoside, 3-*O*- α -L-rhamnopyranosyl-(1 \rightarrow 2)-*O*- α -L-arabinopyranoside, and 3-*O*-sulfate-28-*O*- α -L-rhamnopyranosyl-(1 \rightarrow 4)-*O*- β -D-glucopyranosyl-(1 \rightarrow 6)-*O*- β -D-glucopyranoside; hederagenin 3-*O*- α -L-arabinopyranoside, 3-*O*- α -L-rhamnopyranosyl-(1 \rightarrow 2)-*O*- α -L-arabinopyranoside, 3-*O*- α -L-rhamnopyranosyl-(1 \rightarrow 2)-*O*- α -L-arabinopyranosyl-28-*O*- α -L-rhamnopyranosyl-(1 \rightarrow 4)-*O*- β -D-glucopyranosyl-(1 \rightarrow 6)-*O*- β -D-glucopyranoside, 3-*O*- β -D-glucopyranoside, and 3-*O*- β -D-glucopyranosyl-(1 \rightarrow 2)-*O*- β -D-glucopyranoside; and echinocystic acid 3-*O*- α -L-arabinopyranoside and 3-*O*- α -L-rhamnopyranosyl-(1 \rightarrow 2)-*O*- α -L-arabinopyranoside, in addition to nonglycosylated hederagenin and oleanolic and echinocystic acids.

We used the following solutions: **1-3** (3 g) as the crystalline hydrates (H₇[P(W₂O₇)₆] \cdot *n*H₂O, H₇[P(Mo₂O₇)₆] \cdot *n*H₂O, and H₈[Si(W₂O₇)₆] \cdot *n*H₂O) in CHCl₃:CH₃OH (1:1, v/v, 50 mL); **1-3** (2 g) and *p*-hydroxybenzaldehyde (0.5 g) in CHCl₃:CH₃OH (1:1, v/v, 50 mL); and **1-3** (2 g) in H₂SO₄ (2 N, 50 mL). TLC was performed on Sorbfil plates (RF) grade PTSKh-P-A-UF-254

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of silica-gel particle size 5-7 μm (STKh-1A type sorbent). Chromatograms were heated to 100°C. Elution used $\text{CHCl}_3:\text{CH}_3\text{OH}:\text{NH}_4\text{OH}$ (25%) (100:20:3 and 100:30:5).

Thus, the ability to use reagents containing **1-3** to detect triterpene glycosides and their aglycons on TLC plates was demonstrated. Reagents with **1** and **3** can differentiate glycosides according to the aglycon. Heteropolyacid **3** was used for the first time to detect triterpene glycosides by TLC analysis.

REFERENCES

1. L. A. Yakovishin, *Khim. Prir. Soedin.*, 419 (2003).
2. G. E. Dekanosidze, V. Ya. Chirva, T. V. Sergienko, and N. I. Uvarova, *Investigation of Triterpene Glycosides (Structure Establishment and Synthesis)* [in Russian], Metsniereba, Tbilisi (1982).
3. J. G. Kirchner, *Techniques of Chemistry, Vol. 14: Thin-Layer Chromatography*, 2nd Ed., Wiley-Interscience, New York (1978).
4. R. M. C. Dawson, D. C. Elliott, W. H. Elliott, and K. M. Jones, eds., *Data for Biochemical Research*, Clarendon Press, Oxford (1959).
5. N. I. Grinkevich and L. N. Safronich, eds., *Chemical Analysis of Medicinal Plants* [in Russian], Vysshaya Shkola, Moscow (1983).